

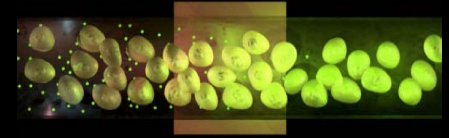
## *MaxCyte STX Workshop*

*Optimizing Cell Based Assays from the Bench to HTS  
Using Scalable Transient Transfection*

*James P. Brady, PhD, MBA*  
*LabAutomation2011, Palm Springs, CA*  
*February 1, 2011*



# Drug Discovery Screening Challenges



Picking the *right target*

Testing therapeutic candidates in a *biologically relevant system*

*scale*

Increasing and decreasing the

of

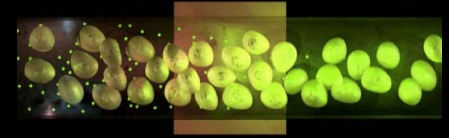
the testing from the bench to HTS to secondary and tertiary screening creates technical, scientific, and process complexity.

*speed*

in

creating and conducting biologically relevant assays is critical to patients and to the financial success of a company.

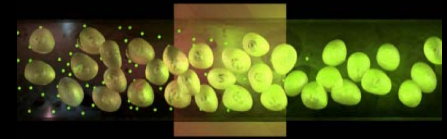
# Ideal Attributes of a Transient Transfection System for Cell Based Assay Development



- Works with multiple cell types
- High efficiency & high viability
- Reproducible from day to day and scientist to scientist
- User friendly
- Loads multiple types of molecules & combinations of molecules
- Compatible with a variety of assay formats

- Allows cryopreservation of transfected cells
- Suitable for multiple classes of targets
- Scalable
- Process compatible
- Short assay development time
- Cost effective

# MaxCyte® STX™ Scalable Transfection System

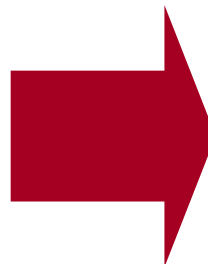


## Proprietary Electroporation Technology



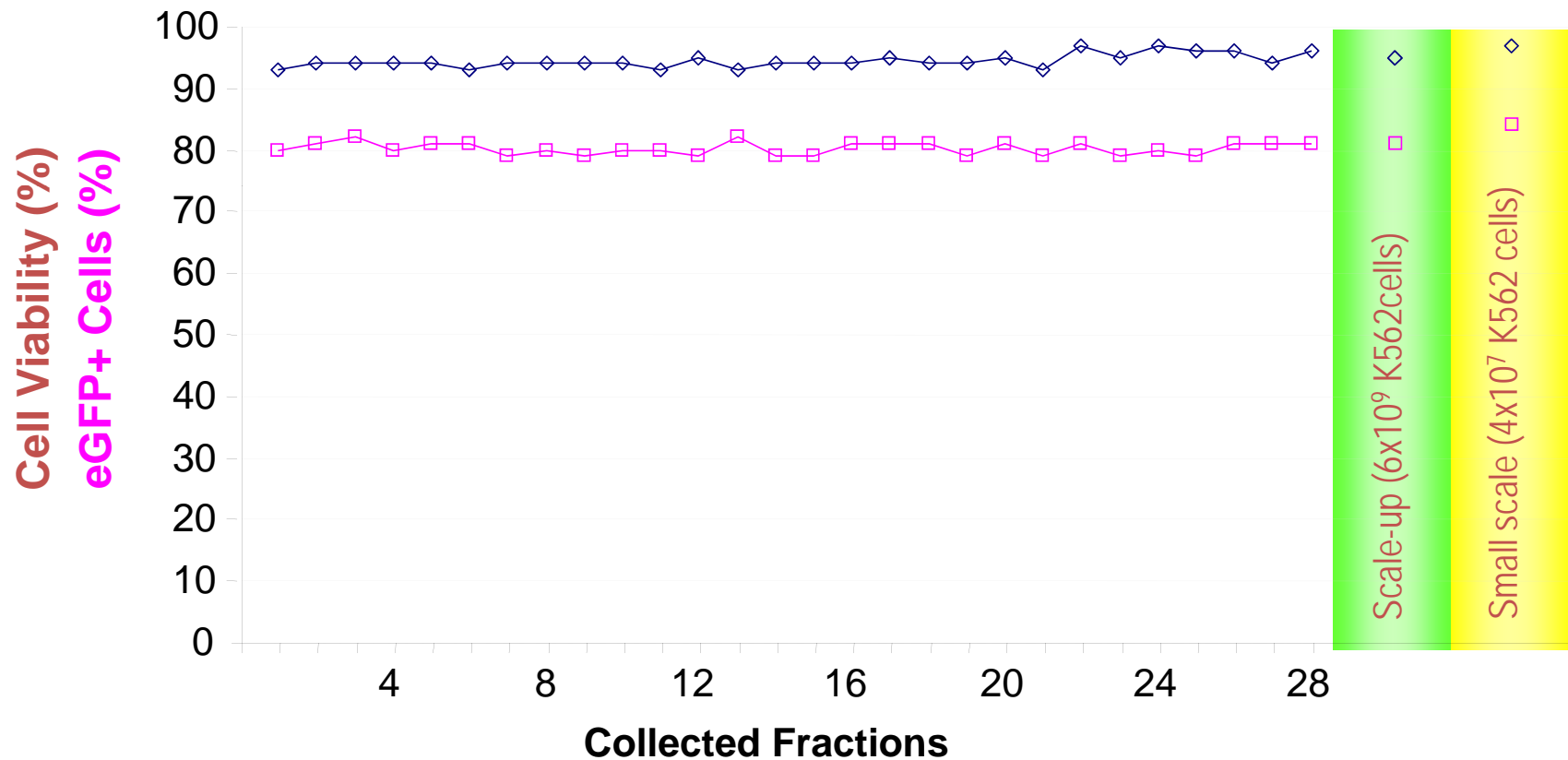
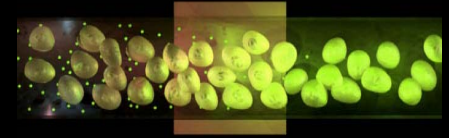
- |                              |                                                                                                                                    |
|------------------------------|------------------------------------------------------------------------------------------------------------------------------------|
| <b>Simple:</b>               | Fast and easy                                                                                                                      |
| <b>High Yield:</b>           | >90% viability                                                                                                                     |
| <b>High Efficiency:</b>      | >90% transfection efficiency                                                                                                       |
| <b>Safe:</b>                 | Chemically defined buffer<br>No added biological agents<br>Sterile, closed system                                                  |
| <b>Scalable &amp; Rapid:</b> | 5 x 10 <sup>5</sup> -4x10 <sup>7</sup> ( <b>Static EP</b> ) in seconds<br>1 x 10 <sup>10</sup> cells ( <b>Flow EP</b> ) in <30 min |
| <b>Rugged:</b>               | Reproducible & consistent                                                                                                          |
| <b>Quality:</b>              | cGMP compliant<br>ISO 9001 certification<br>CE Marking                                                                             |

Small molecules  
Antigens (proteins/lysates)  
Nucleic acids (DNA, mRNA, siRNA)

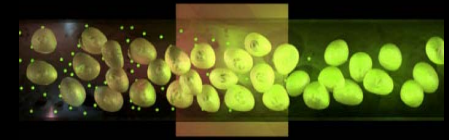


Primary cells  
Stem cells  
Mammalian cell lines

# MaxCyte STX: Reproducible, Consistent, and Scalable



# Representative Sample of MaxCyte STX Transfected Cell Lines & Primary Cells

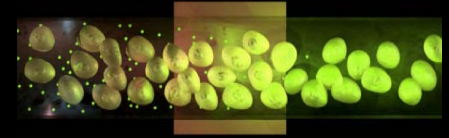


Cell Type	Efficiency % GFP +	Viability
293T	95%	95%
CHO Cells	90%	90%
VERO	90%	90%
K562	90%	90%
NIH 3T3	90%	90%
Jurkat	90%	90%
Huh-7 Cells	80%	90%
Renca	80%	97%
Human Mesenchymal Stem Cells	80%	80%
Human Myoblasts	90%	90%
Human Lymphocytes – B Cells	85%	90%
Human Lymphocytes – T Cells	50%	70%
Human HSC (CD34+ cells)	60%	60%
Human Dendritic Cells	50%	80%

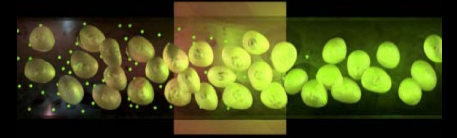
Results of processing cell lines with DNA plasmid encoding for Green Fluorescent Protein (GFP). Efficiency expressed as % cells GFP+ at 24 to 48 hrs post process; viability as % cells excluding propidium iodide (PI).



# Current (and Expanding) MaxCyte STX Protocol List

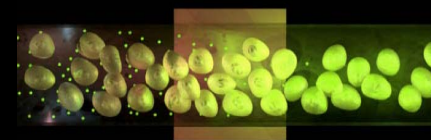


- CHO
- CHO2 (Protein Expression)
- Hela
- HEK 293
- HEK2 (Protein Expression)
- Huh-7
- Jurkat
- K562
- Mesenchymal Stem Cells
- NIH 3T3
- Primary Fibroblasts
- Renca
- Vero
- PC12
- Hep G2
- CV-1
- THP-1
- Min-6
- Panc-1
- L5278Y
- U2OS
- SH-SY5Y
- COS-1
- A549
- PC-3
- BHK-21
- RBL
- Neuro2a
- NS0
- C6
- CaCo-2
- RLE
- COS-7
- LNCaP
- DLD-1
- C2C12



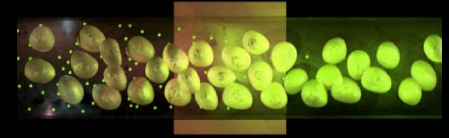
**Developing a Cell Based Assay  
with the  
MaxCyte<sup>®</sup> STX<sup>™</sup>  
Scalable Transfection System**

# How to Use the MaxCyte STX: Assay Development Steps



1. DNA Preparation
2. Cell Culture
3. Electroporation and DNA Titration
4. Cell Handling Post-EP
5. Plating and Analysis
6. Scale-Up
7. Cryopreservation

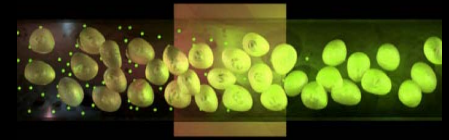
## 1) DNA Preparation: Specifications



- Prepare the DNA from commercially available ion exchange columns, preferably endotoxin-free
- Suspend the DNA in water, not TE
- $OD_{260/280} > 1.8$
- >85% covalently closed, circular conformation
- Stock solution at >5 mg/mL

**High quality DNA is mandatory.**

## 2) Cell Culture: Specifications



- Cells must be in log phase growth
- Typically split the cells one day prior to EP
- Adherent cells must be subconfluent
- Cells should be passaged several times post-thaw
- Late passage cells give poor results
- Cells must be dissociated – no clumps

**Healthy cells are required.**

## 4) Cell Handling Post-EP:



- Immediately remove the cells from the PAs

DO NOT rinse the PAs! DO NOT add media!

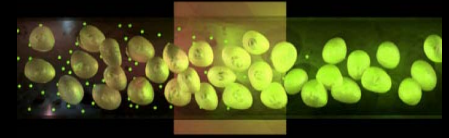
- Cell recovery at 37°C for 20 minutes

Spread cells out for oxygen transfer + better viability.

- Remove the cells and plate as desired.

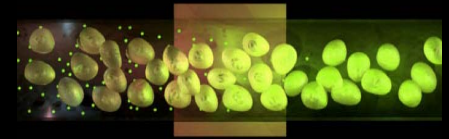
Note: Higher expression levels for ion channels may be obtained by subsequent incubation at 28°C.

## 5) Plating and Analysis:



- Handle cells per standard procedures:  
Plate in multi-well plates or assay chambers
- Assay responsiveness
  - DNA concentration (MaxCyte STX)
  - Number of cells/well
  - Length of time between transfection and analysis

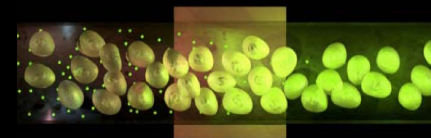
# Seamless Scale-Up: Flow (Large Scale) Electroporation



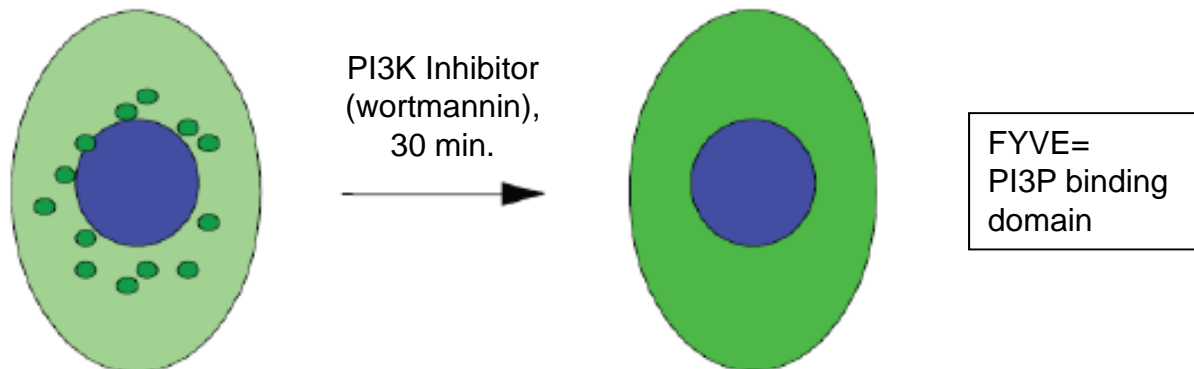
- Choose DNA concentration from the static EP (small scale) experiments
- Prepare appropriate quantity of DNA and cells
- Choose CL-2 from the dropdown menu
- Press start
- < 30 minutes later, recover the cells, plate/cryopreserve, run the assay



# Case Study: Using the MaxCyte STX for Assay Development

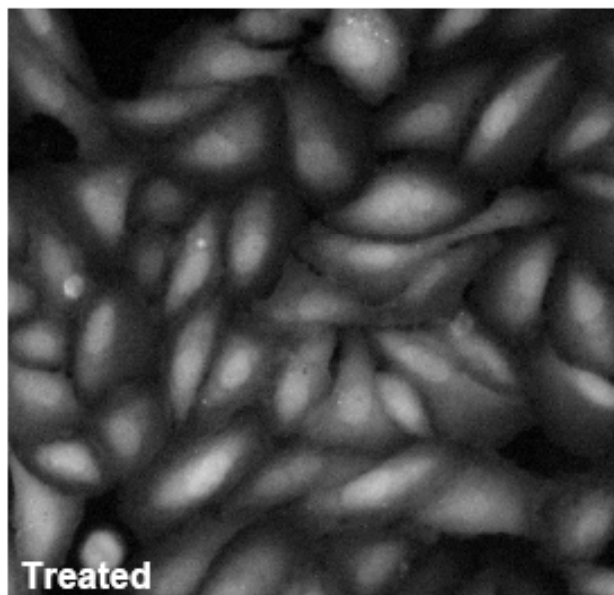
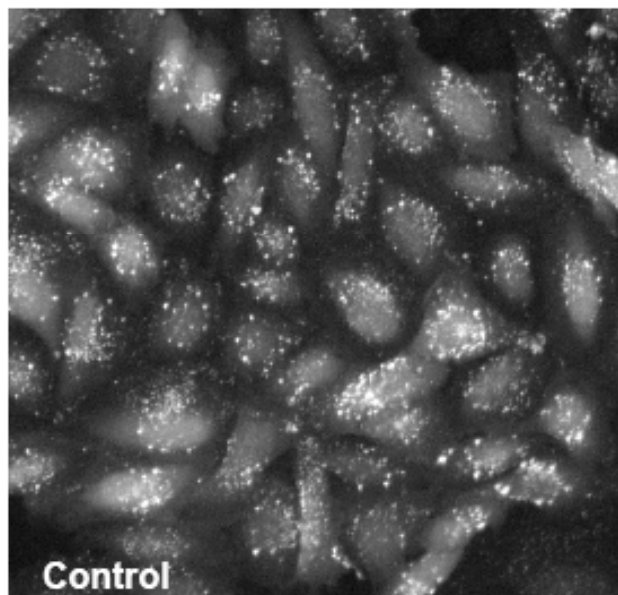


## 2XFYVE-eGFP Redistribution Assay in Tumor Cells: Measuring PI3K Activity



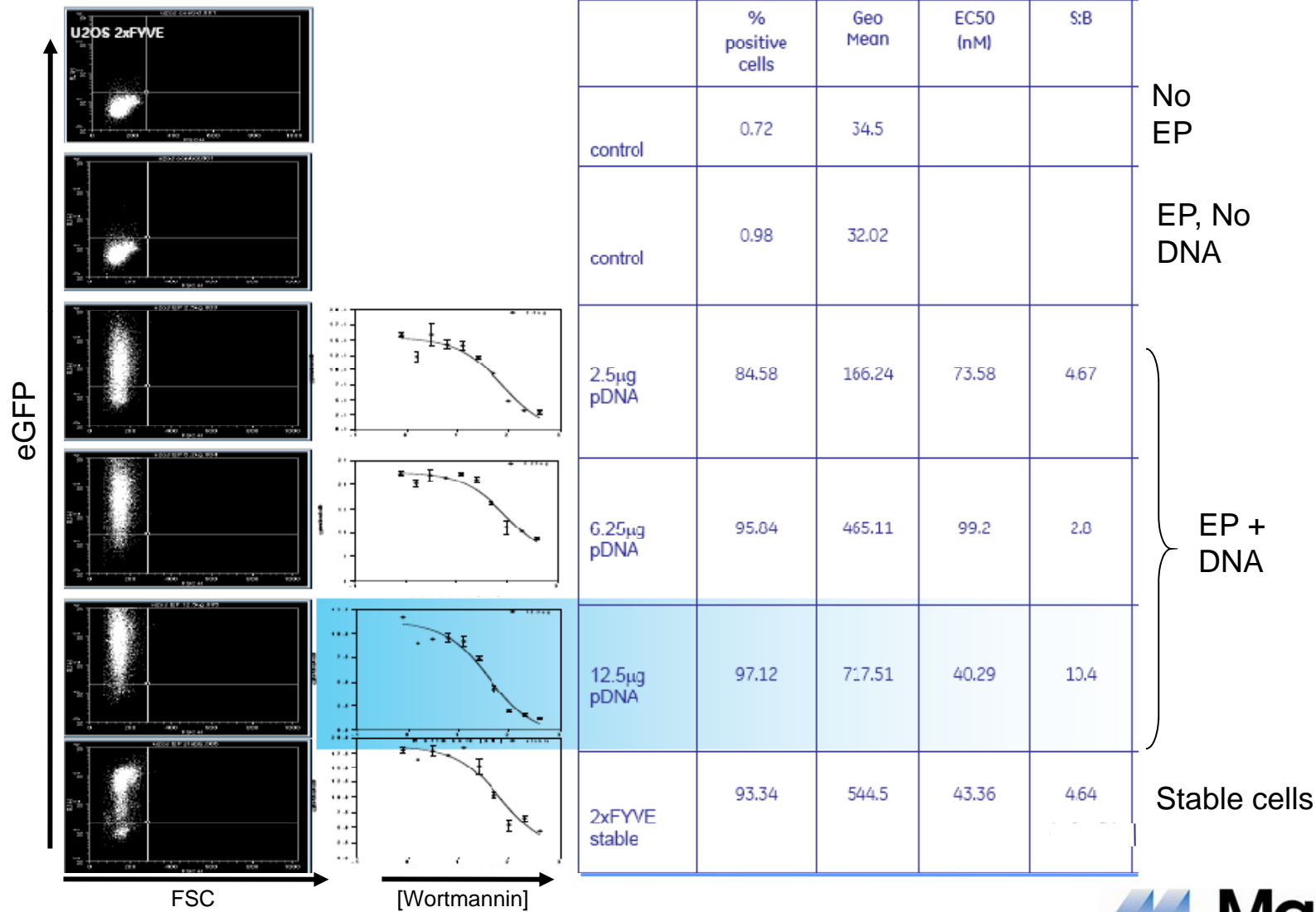
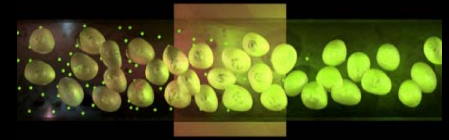
Untreated cell: eGFP-2XFYVE is concentrated in endosomes

Treated cell: eGFP-2XFYVE is redistributed to the cytoplasm

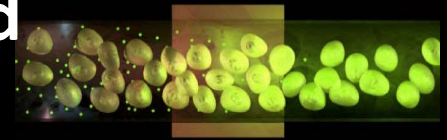


96 Well Assay Format

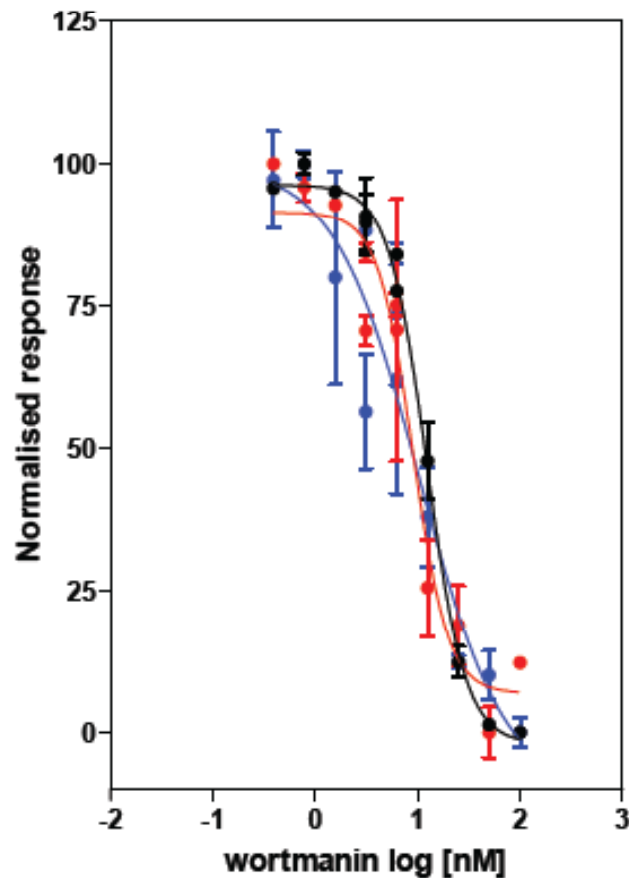
# Optimizing Plasmid DNA Concentrations in Small Scale Transfections



# Seamless Scale-Up: MaxCyte STX-Transfected Cells Comparable to Stable Cell Line

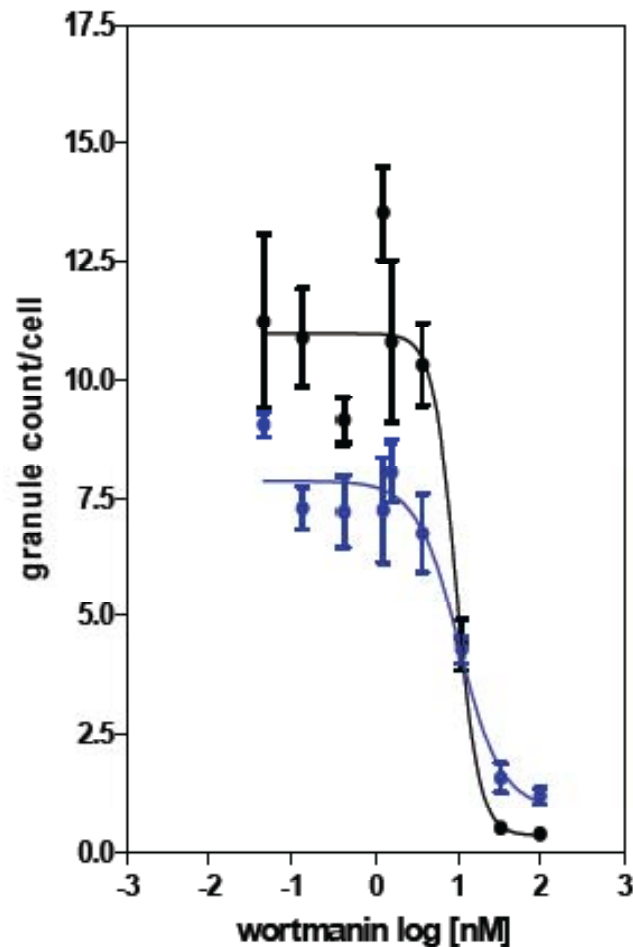
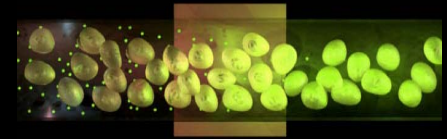


Normalised data



Sample	EC50 (nM)
Stable cell line	12.52
SCEP	10.60
LSEP	9.0

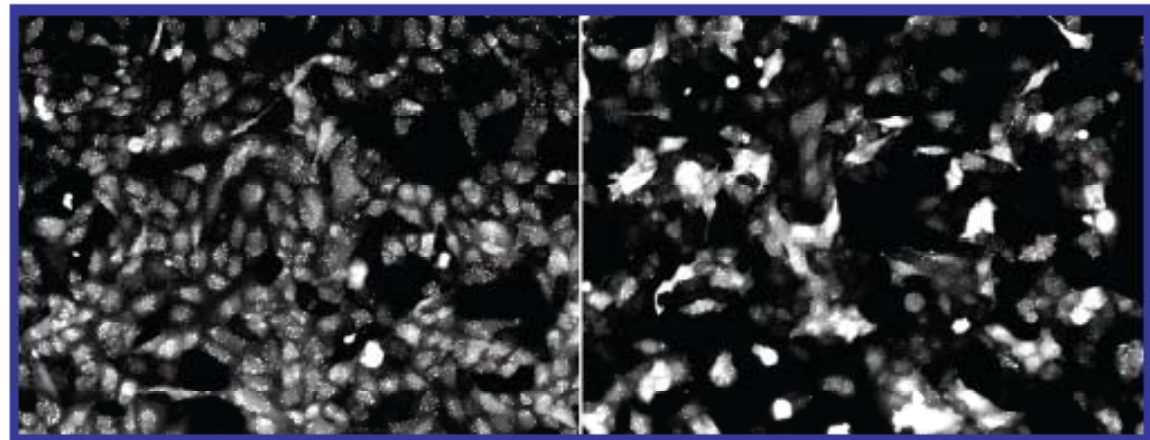
# Assay Results: Cryopreserved Cells Comparable to Stable Cell Line



EP'ed cells seeded into  
4xT162 for 24h  
Cells harvested and 12 x vials  
cryopreserved @ 2E06/ml in  
Medium/FBS/DMSO

T=1 week at -140°C

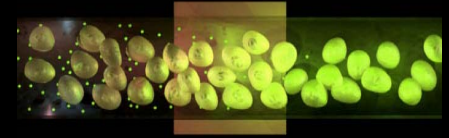
Sample	EC50 (nM)
Stable cell line	10.29
SCEP	9.50



Stable

Control

# MaxCyte STX Advantages



- Small to large volume scalability, ~10 billion cells in < 30 minutes
- Consistent and reproducible transfections
- High cell viability and transfection efficiency
- Transfection of multiple agents simultaneously
- Minimal off-target effects



- More relevant assays
- Faster experimental turnaround
- Greater productivity

**Better Drug Candidates**